## Root physiology – from gene to function

## Preface

### Introduction

In the last decade, enormous progress has been made on the physiology of plant roots, including on a wide range of molecular aspects. Much of that progress has been captured in the following papers, which highlight that the classical boundaries between physiology, biochemistry and molecular biology have vanished. Breakthroughs have been made possible through integration of molecular and whole-plant aspects. There has been a strong focus on a limited number of model species, including Arabidopsis thaliana. That focus has allowed greater insight into the significance of specific genes for plant development and functioning. However, many species are very different from A. thaliana, in that they are mycorrhizal, develop a symbiosis with N2-fixing microsymbionts, or have other specialised root structures. Also, some have a much greater capacity to resist extreme environments due to specific adaptations, for example aerenchyma in the case of soil flooding, or are capable of tolerating a wide range of soil chemical constraints, such as acidity, salinity or heavy metal toxicities. Research on species other than A. thaliana is therefore pivotal, to develop new knowledge in plant sciences in a comprehensive manner. This fundamental new knowledge can be the basis for important applications in, e.g., agriculture and plant conservation. Although significant progress has been made, much remains to be learnt, especially for many aspects of root physiology. It is envisaged, however, that discoveries made in the recent past will likely lead to major breakthroughs in the next decade.

#### Resource acquisition and carbon metabolism

The discovery of the role of aquaporins in water transport in both animals and plants has been a

major breakthrough for understanding plant water relations, in particular water uptake (Vandeleur et al., 2005). Aquaporins are water-channel proteins (Johansson et al., 2000; Maurel, 1997). Their name is somewhat unfortunate, since the term 'porin' is commonly used for proteins that allow the transport of large molecules in a fairly unspecific manner, whereas we now know that aquaporins, which belong to a class of proteins termed MIPs (membrane-intrinsic proteins), are rather specific. However, some can also transport other small molecules, e.g., glycerol (Zeuthen and Klaerke, 1999) or  $CO_2$  (Uehlein et al., 2003). Knowledge on the regulation of aquaporins contributes to understanding plant responses to some abiotic stresses and might help in the design of new irrigation techniques to improve use of scarce water resources. Aquaporins are involved in the adjustment of the hydraulic conductivity of roots, and therefore in adjustments of the roots' physiology during both phases of wet/dry cycles in natural communities and in horticultural crops with irrigation management using partial root-zone drying (Vandeleur et al., 2005). Wider implications of the regulation of aquaporins for acclimation during abiotic stress include the recent finding that the well-known decrease in root hydraulic conductance during flooding involves gating of a root aquaporin, due to cytosolic acidosis (Tournaire-Roux et al., 2003). Moreover, it can be speculated that aquaporins are important in hydraulic redistribution of water in the root-soil system. Aquaporins would allow a plant control over the exit of water from its roots into a soil with a more negative water potential than that of the roots themselves. This phenomenon, which was first described by Caldwell and Richards (1989) and termed 'hydraulic lift', is quite common in desert species with access to deep water (Yoder and Novak, 1999) and re-hydrates the rhizosphere of surface roots during the night. In contrast to 'hydraulic lift', for trees growing in a soil with a low conductance between the top and deeper layers, water movement *down* the profile might rely on hydraulic redistribution via the taproot (Burgess et al., 1998). So far, there are no data to support any speculations on involvement of aquaporins in hydraulic re-distribution, but it is expected that such information will become available during the next decade.

Major progress has been made on the physiology of uptake and metabolism of nitrogen (Miller and Cramer, 2005) and phosphorus (Raghothama and Karthikeyan, 2005; Smith, 2003a). Genes involved in the transport of these nutrients have been identified, and we are beginning to learn when and where these are expressed. We have also gained a much better understanding of the functioning of specialised roots involved in nutrient acquisition, such as the "proteoid" or "cluster" roots of Proteaceae and Lupinus albus (Lambers et al., 2003; Shane and Lambers, 2005). These cluster roots produce and exude vast amounts of carboxylates. Carboxylate release is not restricted to species with cluster roots, but it is the combination of their capacity to release carboxylates in an exudative burst and their structure that allows build-up of high concentrations in the rhizosphere. This ability provides these species with an opportunity to substantially modify their rhizosphere and 'mine' the soil in their immediate vicinity; they are particularly effective in the acquisition of scarcely available phosphorus and micronutrients. Some species belonging to the Cyperaceae have 'dauciform' (=carrot-shaped) roots (Davies et al., 1973; Lamont, 1974; Shane et al., 2005). In many ways, these dauciform roots function in much the same way as cluster roots; their development is suppressed at a high phosphorus supply and when formed they release citrate in an exudative burst (Shane et al., 2005; M.W. Shane and H. Lambers, unpubl.). The combination of biochemical/physiological and anatomical/morphological traits, which allows the build-up of high concentrations of exudates in the rhizosphere might be further exploited considering that the world's phosphorus reserves are dwindling (Vance, 2001).

Major phosphorus reserves are stored in agricultural soils, due to the application of phosphorus fertilisers ('phosphorus bank'). Depending on soil type and agronomic practices in the past, over 80% of applied phosphorus in fertilisers can be sorbed to soil particles, and hence, largely unavailable for most crop plants. We will need crop species with the root traits of Proteaceae and Cyperaceae to access that phosphorus bank. So far, only a limited number of species with cluster roots have been used in agriculture and horticulture, including Macadamia integrifolia (to produce nuts) (Firth et al., 2003), Lupinus albus (as a source of protein) (Gardner et al., 1981) and Aspalanthus linearis (to produce rooibos tea) (Ratshitaka et al., 2001). Most Lupinus species do not make cluster roots of the type produced by L. albus, but they do make structures of a similar function (Hocking and Jeffery, 2004). Other Lupinus species that are used as crop species, e.g., L. angustifolius make sand-binding roots (pers. observation), which may well serve a similar purpose. We are not aware of any species with dauciform roots being intentionally used in managed systems, although, given the relatively wide occurrence of dauciform roots in Cyperaceae (Shane et al., 2005) species with this root type might have been planted in some constructed wetlands. Making greater use of species with cluster roots or similar traits will be of economic benefit, since these plants will be able to access sparingly available phosphorus. In addition, there may be ecological benefits, because an improved capacity for phosphorus acquisition will decrease the need for high rates of phosphorus application, and hence potentially reduce run-off of phosphorus into streams and estuaries, thus limiting eutrophication.

Plants often acquire limiting resources in symbiosis with micro-organisms (Vessey et al., 2005). Our understanding of the legume-rhizobium symbiosis has increased dramatically, not in the least because of the powerful molecular techniques of transcriptomics and metabolomics. These approaches have made it possible to obtain a much improved overview of the metabolic differentiation during nodule development in Lotus japonicus (Colebatch et al., 2004). Approximately 860 genes are more highly expressed in nodules than in roots, including one third involved in metabolism and transport. More than 100 of the highly expressed genes encode proteins likely involved in signalling, or regulation of gene expression at the transcriptional or post-transcriptional level. The analysis showed clear signs of hypoxia in root nodules, as expected; in addition, there were numerous indications that nodule cells also experience phosphorus limitation. Much less is known about other  $N_2$ -fixing symbiosis (Vessey et al., 2005).

Mycorrhizal associations can be found in most vascular plant species, and this field of research has developed enormously in the recent past (Graham and Miller, 2005). Molecular tools have revolutionised studies of mycorrhizal diversity and abundance, improving knowledge on host specificity of the symbionts (Graham and Miller, 2005) and highlighting relationships between mycorrhizal fungi diversity as influencing the structure of some communities of vascular plants (e.g., Reynolds et al., 2003). Like the recent discoveries of genes for transport of mineral nutrients in plants (Miller and Cramer, 2005), rapid advances to elucidate genes regulating exchanges of molecules between plant hosts and arbuscular mycorrhizal (AM) fungi are being made. Our views on 'mycorrhizal dependency' need to be revisited, with the discovery that the micro-symbionts can be responsible for most of the phosphorus uptake when there is no, or only a minor, growth response (Smith et al., 2003b).

Carnivory of small animals is a specialised strategy used by some plant species to acquire inorganic nutrients (Adlassnig et al., 2005). Carnivorous species inhabit a range of nutrientimpoverished environments, ranging from fireprone sand plains to acid peat bogs and aquatic habitats, with, as one might expect, a wide range of root sizes and structures (Adlassnig et al., 2005). In contrast with information available on their traps formed by shoot organs (Juniper et al., 1989), comparatively little is known about the physiology of roots of carnivorous plants; several studies have demonstrated the importance of the roots in water and nutrient uptake for at least some carnivorous species, but not in several others.

Since two books (Day et al., 2004; Lambers and Ribas-Carbo, 2005) dealing with plant respiration have recently been published in Govindjee's series *Advances in Photosynthesis and Respiration*, no review dealing with this aspect of root physiology has been included. Over the last decade or so, we have gained a much better understanding of the respiratory costs of nutrient acquisition in fast- and slow-growing herbaceous species (Poorter et al., 1991; Scheurwater et al., 1998). When plants are grown at a high supply of nitrate, the costs associated with nitrate uptake are considerably greater for inherently slow-growing species, due to a major efflux component (Scheurwater et al., 1999). That passive efflux needs to be compensated by a greater active influx, and thus accounts for greater respiratory costs per unit N acquired. However, greater efflux is not the cause of slower growth; rather, slower growth leads to greater efflux (Ter Steege et al., 1999) and hence greater respiratory costs (Nagel and Lambers, 2002). Considerable information has also become available on the quantitative significance of cyanide-resistant root respiration (Millenaar et al., 2001). Major progress is to be expected on the physiological significance of the alternative path, which has puzzled plant scientists for quite some time. That progress can be expected, because a technique to assess the activity of this path in intact tissues has become more widely available (Ribas-Carbo et al., 2005).

### Perception of the abiotic and biotic root environment

We have gained an appreciation of the fact that roots 'sense' their environment, and that the plant responds in a feed-forward and adaptive manner. The feed-forward response when roots sense adverse conditions in the soil allows acclimation before a major disturbance of the plant's metabolism (Davies et al., 1994). When roots perceive that the soil is flooded, they respond with an enhanced concentration of ethylene in both the roots and above-ground plant parts. Ethylene induces the formation of aerenchyma in roots (Visser and Voesenek, 2005), and also affects adaptive processes in above-ground organs (Voesenek and Blom, 1999). Similarly, roots can sense the availability of water (Davies et al., 1994) as well as nitrogen and phosphorus (De Groot et al., 2003), and signal this information to the shoot, which responds in a feed-forward manner. In the case of water stress, the signalling molecule is ABA (Davies et al., 1994; Schurr et al., 1992). Cytokinins are involved in signalling the plant's N and P status (De Groot et al., 2003; Kuiper et al., 1989; Lambers et al., 1998). These are some of many examples showing

'signalling' between roots and shoots. More details about signals and signal-transduction pathways are included in the review by Dodd (2005), highlighting that the signals need not always be one of the classical phytohormones, but can, for example, be nitrate or sugars also. Internal signalling in plants, as dependent on plant development and environmental conditions is a rapidly developing field, where much progress can be expected. The challenge will be to integrate the new information into improved crop production systems.

Sensing does not only involve resources, but also other chemical factors in the environment. Aluminium-resistant genotypes must be able to sense the presence of aluminium to respond with the release of aluminium-complexing carboxylates (Kochian et al., 2005). When roots release either mainly citrate or mainly malate, depending on soil pH (Veneklaas et al., 2003), they must be sensing the soil pH or a factor closely associated with it. Sensing environmental conditions is obviously crucial to performance of plants, although we still lack a thorough understanding of the exact mechanisms involved.

Signalling in plants is not only important between different organs of a plant, but also between hosts and parasitic plants (Okubara and Paulitz, 2005). Holoparasitic species are entirely dependent on a host for the completion of their life cycle (Lambers et al., 1998). If their seeds were to germinate in the absence of a suitable host, that would be fatal, especially for those that have extremely small seeds. In fact, both germination (Siame et al., 1993) and the formation of haustoria (Estabrook and Yoder, 1998; Smith et al., 1990), which connect the parasite with its host, depend on chemical signalling between host and parasite. This is an exciting and rapidly developing field. Knowledge of these interactions may appear esoteric, but major applications can be expected, because some parasitic species (e.g., Striga and Orobanche species) belong to the world's worst weeds (Emechebe et al., 2004; Marley et al., 2004; Rodriguez-Conde et al., 2004). Others (Cistanche species) are grown to produce medicine in north-eastern China (Geng et al., 2004). Low-cost and safe signalling molecules that trigger the germination of the seeds of parasitic pest species before crops are sown might be of enormous benefit, especially to farmers in developing countries.

Signalling is also important between hosts and their symbiotic micro-organisms, e.g., rhizobia (Vessey et al., 2005) and mycorrhizal fungi (Graham and Miller, 2005). The intricate interactions that precede the establishment of a functional symbiosis are best understood for the rhizobium– legume symbiosis (Esseling and Emons, 2004). They must also play a role in other symbiotic systems that fix dinitrogen, but the progress in that area has been much slower (Rai et al., 2000; Vessey et al., 2005). Somewhat more is known on signalling between hosts and some mycorrhizal fungi (Graham and Miller, 2005), but much remains to be discovered.

# Plants growing in soil with adverse abiotic or biotic conditions

Some species or ecotypes are capable of growing in soils that are naturally enriched with heavy metals, e.g., serpentine or ultramafic soils. The metals may be 'excluded' or absorbed, and stored in compartments where they do not harm the plant's metabolism (Meharg, 2005). Recently, progress has been made on the mechanisms accounting for internal transport and storage of heavy metals as well as metal 'exclusion' (Meharg, 2005). Species or ecotypes that accumulate heavy metals to very high levels are called metallophytes. These are not restricted to soils naturally enriched with heavy metals, but are also found on sites contaminated by heavy metals, e.g., due to mining. Such metallophytes are very important to stabilise contaminated soil, and stop it from spreading over a larger area. Metallophytes have been proposed as a method to clean contaminated soil (phytoremediation) (Meharg, 2005) or extract metals from soil with the intention to mine the metals (phytomining) (Li et al., 2003). To be economically viable options, the metallophytes have to accumulate metals to very high concentrations and produce a lot of biomass in a relatively short time. However, most metallophytes are inherently slow-growing, most likely because they were selected in lownutrient environments, which are typically inhabited by slow-growing species (Lambers and Poorter, 1992). Genotypes that are both metal resistant and productive need to be selected. Major new discoveries are to be expected in the next

decade, but applications in the context of phytoremediation and phytomining would appear to be less promising than claimed when first proposed, unless combined with other profit-making operations, e.g., forestry (Robinson et al., 2003).

Mycorrhizas have been claimed to 'protect' higher plants from negative effects of heavy metals in soil (Leyval et al., 1997). It has also been shown that species that belong to a typically nonmycorrhizal family can be mycorrhizal if they are associated with soils with high levels of heavy metals. One example is for California serpentine grassland communities, where Arenaria douglasii (Caryophyllaceae) and Streptanthus glandulosus (Brassicaceae) were found to be mycorrhizal (Hopkins, 1987). Another example is for Hakea verrucosa (Proteaceae), occurring on nickel-containing ultramafic soils in Western Australia (Boulet and Lambers, 2005). These are exciting observations, from an evolutionary perspective as well as because of the possible implications for the rehabilitation of contaminated sites after mining.

Acid soils represent another stress to plant roots. It is not so much the low pH itself that causes the problems, but the fact that the solubility of specific metals strongly depends on pH (Kochian et al., 2005). In particular aluminium is considerably more soluble at low pH. Acid-resistant species typically are aluminium resistant. Resistance is at least partially based on 'exclusion', due to precipitation with citrate, malate or oxalate released from roots, depending on the species (Delhaize et al., 1993; Zheng et al., 1998). Some aluminium-resistant species also accumulate aluminium, and detoxify it internally as aluminium-carboxylate precipitates (Ma et al., 1997; Zheng et al., 1998). As for heavy metal resistance (Meharg, 2005), there is considerable genetic variation for aluminium exclusion (Delhaize et al., 1993; Kochian et al., 2005). This allows important application in agriculture, but care also has to be taken to minimise further soil acidification, which is a common trend in cropping systems (Lambers et al., 1998). Soil amelioration, e.g., using lime, remains important, but does not invariably address sub-soil acidity; therefore, aluminium-resistant genotypes would be important to develop.

Drought, salinity and flooding are abiotic stresses each of which influence plant species composition and productivity in natural and managed systems. Improved knowledge on physiological responses of roots to water stress, including root-to-shoot signalling (Dodd, 2005) should aid design of new irrigation techniques to improve use of scarce water resources (Vandeleur et al., 2005). Since various aspects of resistance to salinity have been reviewed recently (e.g., Munns, 2002; Tester and Davenport, 2003), and in a special issue dealing with dryland salinity in Australia (Lambers, 2003), salinity is not covered in this volume. Flooding regimes shape the composition of natural communities in some areas (Voesenek et al., 2004) and underpin rice production systems; soil waterlogging is also a problem in vast areas of irrigated and rainfed agriculture. Visser and Voesenek (2005) provide a comprehensive review on signals, and signal-transduction pathways, crucial to the perception and acclimation by plant roots to soil flooding. Hormones and signalling pathways that regulate traits for flooding resistance, such as adventitious rooting, aerenchyma formation for gas transport (Colmer and Greenway, 2005; Jackson and Armstrong, 1999) and root metabolism during O<sub>2</sub> deficiency (Gibbs and Greenway, 2003; Jackson and Ricard, 2003) are reviewed by Visser and Voesenek (2005). Substantial gains in knowledge in some areas are highlighted, as are emerging topics that are still poorly understood and will be priority areas for future research.

Roots frequently encounter adverse biotic conditions, due to the presence of microbial pathogens, nematodes, viruses and plant parasites (Okubara and Paulitz, 2005). Our understanding of these interactions has increased enormously, again, in part due to the development of new molecular tools. Developing resistance to root pests and diseases will continue to be important, as chemical protection is not always a desirable option.

# Using new genotypes and combinations of crop species based on new ecophysiological information

Allelopathic interactions are very hard to demonstrate in nature, but they are very likely to occur, also in managed ecosystems (Lambers et al., 1998). The interactions may involve micro-organisms (Inderjit, 2005). Allelopathic interactions may account for the invasive nature of some weeds (Ridenour and Callaway, 2001). There are major possibilities for applications in agriculture. Accessions of wheat (*Triticum aestivun*) differ widely in their potential to inhibit seed germination of ryegrass (*Lolium rigidumi*) (Wu et al., 2000a), a major weed in Australia (Powles and Shaner, 2001). That variation appears to be associated with the release of allelochemicals of a phenolic nature (Wu et al., 2000b), although so far the phenolics have only been assessed in root tissue, not in exudates. Making a crop more competitive, by enhancing its capacity for interference competition, would reduce the need for herbicides.

Facilitation refers to positive effects of one plant on another (Callaway, 1995). It is equally difficult to demonstrate in natural systems as allelopathic interactions are (Hauggaard-Nielsen and Jensen, 2005), but there are numerous examples of increased yields when combinations of crop species are used (Karpenstein-Machan and Stuelpnagel, 2000; Zuo et al., 2000). Such agronomic practices, usually called intercropping, are used in the low-input systems of the tropics, where crops are harvested manually (Willey, 1979), and are also common in China (Zhang et al., 2004) where new intercropping systems continue to be developed (Guixin et al., 2004). If reliable systems can be developed to mechanically harvest intercropped species at the same time, then this would stimulate the development of the practice for broad-area agriculture in other countries.

Pasture agronomy already uses combinations of species in broad-area agriculture, but further research might enhance productivity of pasture systems if the best combinations of species, and perhaps genotypes within species, can be further refined. For example, when seedlings of mycorrhizal and non-mycorrhizal species are grown together, they tend to have negative effects on each other which are not seen when either seedlings of mycorrhizal species or ones of non-mycorrhizal species grow together (Francis and Read, 1994). The chemical basis of this interference is not known, but the observation may have major implications for plant functioning in natural or managed systems. It may mean that combinations of mycorrhizal and non-mycorrhizal species are less desirable for intercropping, and this will need to be addressed to enhance productivity of intercropping and pasture systems.

#### Perspectives

Many new discoveries are to be expected in the ecophysiology of roots of native and crop species. One can envisage many applications of the new fundamental knowledge. One area that has not been reviewed in this volume, because it is too new to have generated many publications, is that of signalling in tritrophic below-ground interactions. Similar above-ground interactions are well documented for interactions between plants, their herbivores and 'bodyguards' (Alborn et al., 1997; Kessler and Baldwin, 2001; Sabelis et al., 2001). The first exciting information is now becoming available on interactions between roots of Thuja occidentalis, which release chemicals upon attack by weevil larvae (Otiorhynchus sulcatus), to attract parasitic nematodes (Heterorhabditis megidis), which then prey on the weevil larvae (Van Tol et al., 2001). Similar tritrophic interactions appear to occur in Zea mays (T.C.J. Turlings, pers. comm.). It is to be expected that improved knowledge in this area should provide opportunities for applications in plant management systems, similar to those existing for aboveground tritrophic interactions (Turlings and Wäckers, 2004).

Major progress on understanding numerous aspects of root physiology, and under several important environmental constraints, has been made possible by close interactions between ecophysiologists, biochemists and molecular geneticists. These close interactions will be important to achieve new breakthroughs in yield improvement. Such breakthroughs are vitally important, if we are to produce enough food and fibre for the worlds growing population in a sustainable manner.

#### References

- Adlassnig W, Peroutka M, Lambers H and Lichtscheidl I K 2005 The roots of carnivorous plants. Plant Soil 274, 127–140.
- Alborn H T, Turlings T C J, Jones T H, Stenhagen G, Loughrin J H and Tumlinson J H 1997 An elicitor of plant volatiles from beet armyworm oral secretion. Science 276, 945–949.
- Boulet F M and Lambers H 2005 Characterisation of arbuscular mycorrhizal colonisation in the cluster roots of *Hakea verrucosa* F. Muell (Proteaceae) and its effect on growth and nutrient acquisition in ultramafic soil. Plant Soil 269, 357–367.

- Burgess S S O, Adams M A, Turner N C and Ong C K 1998 The redistribution of soil water by tree root systems. Oecologia 115, 306–311.
- Caldwell M M and Richards J H 1989 Hydraulic lift: water efflux from upper roots improves effectiveness of water uptake by deep roots. Oecologia 79, 1–5.
- Callaway R M 1995 Positive interactions among plants. Bot. Rev. 61, 306–349.
- Chu G X, Shen Q R and Cao J L 2004 Nitrogen fixation and N transfer from peanut to rice cultivated in aerobic soil in an intercropping system and its effect on soil N fertility. Plant Soil 263, 17–27.
- Colebatch G, Desbrosses G, Ott T, Krusell L, Montanari O, Kloska S, Kopka J and Udvardi M K 2004 Global changes in transcription orchestrate metabolic differentiation during symbiotic nitrogen fixation in *Lotus japonicus*. Plant J. 39, 487–512.
- Colmer T D and Greenway H 2005 Oxygen transport, respiration, and anaerobic carbohydrate catabolism in roots in flooded soils. *In Plant Respiration – From Cell to Ecosystem*. Eds. H Lambers and M Ribas-Carbo. Springer, Dordrecht, 137–158.
- Davies J, Briarty L G and Rieley J O 1973 Observations on the swollen lateral roots of the Cyperaceae. New Phytol. 72, 167–174.
- Davies W J, Tardieu F and Trejo C L 1994 How do chemical signals work in plants that grow in drying soil? Plant Physiol. 104, 309–314.
- Day D A, Millar A H and Whelan J 2004 Plant Mitochondria: From Genome to Function. (Advances in Photosynthesis and Respiration, Vol 17). Kluwer Academic Publishers, Dordrecht.
- De Groot C, Marcelis L F M, Van den Boogaard R, Kaiser W M and Lambers H 2003 Interaction of nitrogen and phosphorus nutrition in determining growth. Plant Soil 248, 257–269.
- Delhaize E, Ryan P R and Randall P J 1993 Aluminum tolerance in wheat (*Triticum aestivum* L.). II. Aluminumstimulated excretion of malic acid from root apices. Plant Physiol. 103, 695–702.

Dodd I C 2005 Root-to-shoot signalling. Plant Soil (this volume).

- Emechebe A M, Ellis-Jones J, Schulz S, Chikoye D, Douthwaite B, Kureh I, Tarawali G, Hussaini M A and Kormawa P 2004 Farmers perception of the *Striga* problem and its control in Northern Nigeria. Exp. Agric. 40, 215–232.
- Esseling J J and Emons A M C 2004 Dissection of Nod factor signalling in legumes: cell biology, mutants and pharmacological approaches. J. Microscopy 214, 104–113.
- Estabrook E M and Yoder J I 1998 Plant–plant communication: rhizosphere signaling between parasitic angiosperms and their hosts. Plant Physiol. 116, 1–7.
- Firth D J, Whalley R D B and Johns G G 2003 Distribution and density of the root system of macadamia on krasnozem soil and some effects of legume groundcovers on fibrous root density. Aust. J. Exp. Agric. 43, 503–514.
- Francis R and Read D J 1994 The contribution of mycorrhizal fungi to the determination of plant community structure. Plant Soil 159, 11–25.
- Gardner W K, Parbery D G and Barber D A 1981 Proteoid root morphology and function in *Lupinus albus*. Plant Soil 60, 143–147.
- Geng X C, Song L W, Pu X P and Tu P F 2004 Neuroprotective effects of phenylethanoid glycosides from *Cistanches salsa* against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine

(MPTP) induced dopaminergic toxicity in C57 mice. Biol. Pharmac. Bull. 27, 797–801.

- Gibbs J and Greenway H 2003 Mechanisms of anoxia tolerance in plants. I. Growth, survival and anaerobic catabolism. Funct. Plant Biol. 30, 1–47.
- Graham J H and Miller R M 2005 Mycorrhizas: gene to function. Plant Soil 274, 79–100.
- Guixin C, Qiong S and Jinliu C 2004 Nitrogen fixation and N transfer from peanut to rice cultivated in aerobic soil in an intercropping system and its effect on soil N fertility. Plant Soil (in press).
- Hauggaard-Nielsen H and Jensen E S 2005 Facilitative root interactions in intercrops. Plant Soil 274, 237–250.
- Hocking P and Jeffery S 2004 Cluster root production and organic anion exudation in a group of old world lupins and a new world lupin. Plant Soil 258, 135–150.
- Hopkins N A 1987 Mycorrhizae in a California serpentine grassland community. Can. J. Bot. 65, 484–487.
- Inderjit 2005 Soil microorganisms: an important determinant of allelopathic activity. Plant Soil 274, 227–236.
- Jackson M B and Armstrong W 1999 Formation of aerenchyma and the processes of plant ventilation in relation to soil flooding and submergence. Plant Biol. 1, 274–287.
- Jackson M B and Ricard B 2003 Physiology, biochemistry and molecular biology of plant root systems subjected to flooding of the soil. *In* Root Ecology. Eds. H De Kroon and E J W Visser. pp. 193–213. Springer-Verlag, Berlin.
- Johansson I, Karlsson M, Johanson U, Larsson C and Kjellbom P 2000 The role of aquaporins in cellular and whole plant water balance. Biochim. Biophys. Acta 1465, 324–342.
- Juniper B E, Robins R J and Joel D M 1989 The Carnivorous Plants. Academic Press Limited, London. 353 pp.
- Karpenstein-Machan M and Stuelpnagel R 2000 Biomass yield and nitrogen fixation of legumes monocropped and intercropped with rye and rotation effects on a subsequent maize crop. Plant Soil 218, 215–232.
- Kessler A and Baldwin I T 2001 Defensive function of herbivore-induced plant volatile emissions in nature. Science 291, 2141–2144.
- Kochian L V, Piñeros M A and Owen Hoekenga O A 2005 The physiology, genetics and molecular biology of plant aluminum tolerance and toxicity. Plant Soil, (This Volume).
- Kuiper D, Kuiper P J C, Lambers H, Schuit J T and Staal M 1989 Cytokinin concentration in relation to mineral nutrition and benzyladenine treatment in *Plantago major* ssp. *pleiosperma*. Physiol. Plant. 75, 511–517.
- Lambers H 2003 Introduction, dryland salinity: a key environmental issue in southern Australia. Plant Soil 218, 5–7.
- Lambers H and Poorter H 1992 Inherent variation in growth rate between higher plants: a search for physiological causes and ecological consequences. Adv. Ecol. Res. 22, 187–261.
- Lambers H, Chapin F S III and Pons T L 1998 Plant Physiological Ecology. Springer-Verlag, New York.
- Lambers H, Cramer M D, Shane M W, Wouterlood M, Poot P and Veneklaas E J 2003 Structure and functioning of cluster roots and plant responses to phosphate deficiency. Plant Soil 248, ix-xix.
- Lambers H and Ribas-Carbo M (Eds) 2005 Plant Respiration. From Cell to Ecosystem. (Advances in Photosynthesis and Respiration, Vol 18). Springer, Dordrecht.
- Lamont B 1974 The biology of dauciform roots in the sedge *Cyathochaete avanaceae*. New Phytologist 73, 985–996.

Li Y -M, Chaney R, Brewer E, Roseberg R, Angel J S, Baker A, Reeves R and Nelkin J 2003 Development of a technology for commercial phytoextraction of nickel: economic and technical considerations. Plant Soil 249, 107–115.

- Leyval C, Turnau K and Haselwandter K 1997 Effect of heavy metal pollution on mycorrhizal colonization and function: physiological, ecological and applied aspects. 7, 139–153.
- Ma J F, Hiradata S, Nomoto K, Iwashita T and Matsumoto H 1997 Internal detoxification mechanisms of Al in hydrangea. Identification of Al forms in the leaves. Plant Physiol. 113, 1033–1039.
- Marley P S, Aba D A, Shebayan J A Y, Musa R and Sanni A 2004 Integrated management of *Striga hermonthica* in sorghum using a mycoherbicide and host plant resistance in the Nigerian Sudano-Sahelian savanna. Weed Res. 44, 157–162.
- Maurel C 1997 Aquaporins and water permeability of plant membranes. Annu. Rev. Plant Physiol. Plant Mol. Biol. 48, 399–429.
- Meharg A M 2005 Mechanisms of plant resistance to metal and metalloid ions and potential biotechnological applications. Plant Soil 274, 163–174.
- Millenaar F F, Fiorani F, Gonzalez-Meler M, Welschen R, Ribas-Carbo M, Siedow J N, Wagner A M and Lambers H 2001 Regulation of alternative oxidase activity in six wild monocotyledonous species; an *in vivo* study at the whole root level. Plant Physiol. 126, 376–387.
- Miller A J and Cramer M D 2005 Root nitrogen acquisition and assimilation. Plant Soil 274, 1–36.
- Munns R 2002 Comparative physiology of salt and water stress. Plant Cell Environ. 25, 239–250.
- Nagel O W and Lambers H 2002 Changes in the acquisition and partitioning of carbon and nitrogen in the gibberellindeficient mutants A70 and W335 of tomato (*Solanum lycopersicum* L.). Plant Cell Environ. 25, 883–891.
- Okubara P A and Paulitz T C 2005 Root responses to necrotrophic pathogens and parasites-defense molecules, genes, and pathways. Plant Soil 274, 215–226.
- Poorter H, Van der Werf A, Atkin O and Lambers H 1991 Respiratory energy requirements depend on the potential growth rate of a plant-species. Physiol. Plant. 83, 469–475.
- Powles S B and Shaner D L, (Eds.) 2001 Herbicide Resistance and World Grains. CRC Press, Boca Raton.
- Raghothama K G and Karthikeyan A S 2005 Phosphate acquisition. Plant Soil 274, 37–49.
- Rai A N, Söderback E and Bergman B 2000 Cyanobacteriumplant symbioses. New Phytol. 147, 449–481.
- Ratshitaka T A, Sprent J I and Dakora F D 2001 Nodulated Lupinus albus and Aspalathus linearis and changes in tissue and rhizosphere soil concentrations of P and Al associated with cluster root formation in A. linearis. In Book of Abstracts for a Workshop on Structure and Functioning of Cluster Roots and Plant Responses to Phosphate Deficiency, 2001, Perth, Australia.
- Reynolds H L, Packer A, Bever J D and Clay K 2003 Grassroots ecology: plant–microbe–soil interactions as drivers of plant community structure and dynamics. Ecology 84, 2281–2291.
- Ribas-Carbo M, Robinson S A and Giles L 2005 The application of the oxygen-isotope technique to assess respiratory pathway partitioning. *In* Plant Respiration. From Cell to Ecosystem. Eds. H Lambers and M Ribas-Carbo. Springer, Dordrecht, 31–42.

- Ridenour W M and Callaway R M 2001 The relative importance of allelopathy in interference: the effects of an invasive weed on a native bunchgrass. Oecologia 126, 444–450.
- Robinson B, Fernández J -E, Madejón P, Marañón T, Murillo J M, Green S and Clothier B J 2003 Phytoextraction: an assessment of biogeochemical and economic viability. Plant Soil 249, 117–125.
- Rodriguez-Conde M F, Moreno M T, Cubero J I and Rubiales D 2004 Characterization of the Orobanche–Medicago truncatula association for studying early stages of the parasite– host interaction. Weed Res. 44, 218–223.
- Sabelis M W, Janse A and Kant M R 2001 The enemy of my enemy is my ally. Science 291, 2104–2105.
- Scheurwater I, Cornelissen C, Dictus F, Welschen R and Lambers H 1998 Why do fast- and slow-growing grass species differ so little in their rate of root respiration, considering the large differences in rate of growth and ion uptake? Plant Cell Environ. 21, 995–1005.
- Scheurwater I, Clarkson D T, Purves J, van Rijt G, Saker L, Welschen R and Lambers H 1999 Relatively large nitrate efflux can account for the high specific respiratory costs for nitrate transport in slow-growing grass species. Plant Soil 215, 123–134.
- Schurr U, Gollan T and Schulze E -D 1992 Stomatal response to drying soil in relation to changes in the xylem sap composition of *Helianthus annuus*. II. Stomatal sensitivity to abscisic acid imported from the xylem sap. Plant Cell Environ. 15, 561–567.
- Shane M W and Lambers H 2005 Cluster roots: a curiosity in context. Plant Soil 274, 101–125.
- Shane M W, Dixon K W and Lambers H 2005 An investigation of the occurrence of dauciform roots amongst Western Australian reeds, rushes and sedges, and the impact of P supply on dauciform-root development in *Schoenus unispiculatus* (Cyperaceae). New Phytol. 165, 887–888.
- Siame B P, Weerasuriya Y, Wood K, Ejeta G and Butler L G 1993 Isolation of strigol, a germination stimulant for *Striga* asiatica, from host plants. J. Agric. Food Chem. 41, 1486– 1491.
- Smith C E, Dudley M W and Lynn D G 1990 Vegetative/ parasitic transition: control and plasticity in *Striga* development. Plant Physiol. 93, 208–215.
- Smith F W, Mudge S R, Rae A L and Glassop D 2003a Phosphate transport in plants. Plant Soil 248, 71–83.
- Smith S E, Smith F A and Jakobsen I 2003b Mycorrhizal fungi can dominate phosphate supply to plants irrespective of growth responses. Plant Physiol. 133, 18–20.
- Ter Steege M W, Stulen I, Wiersema P K, Posthumus F and Vaalburg W 1999 Efficiency of nitrate uptake in spinach: impact of external nitrate concentration and relative growth rate on nitrate influx and efflux. Plant Soil 208, 125–134.
- Tester M and Davenport R 2003 Na<sup>+</sup> tolerance and Na<sup>+</sup> transport in higher plants. Ann. Bot. 91, 503–527.
- Tournaire-Roux C, Sutka M, Javot H, Gout E, Gerbeau P, Luu D -L, Bligny R and Maurel C 2003 Cytosolic pH regulates root water transport during anoxic stress through gating of aquaporins. Nature 425, 393–397.
- Turlings T C J and Wäckers F L 2004 Recruitment of predators and parasitoids by herbivore-damaged plants. *In* Advances in Insect Chemical Ecology. Eds. R T Cardé and J Millar. pp. 21–75. Cambridge University Press, Cambridge.

- Uehlein N, Lovisolo C, Siefritz F and Kaldenhoff R 2003 The tobacco aquaporin tAQP1 is a membrane CO<sub>2</sub> pore with physiological functions. Nature 425, 734–737.
- Vance C P 2001 Symbiotic nitrogen fixation and phosphorus acquisition. Plant nutrition in a world of declining renewable resources. Plant Physiol. 127, 390–397.
- Vandeleur R, Niemietz C and Tyerman S D 2005 Roles of aquaporins in responses to irrigation. Plant Soil 274, 141–161.
- Van Tol R W H M, Van der Sommen A T C, Boff M I C, Van Bezooijen J, Sabelis M W and Smits P H 2001 Plants protect their roots by alerting the enemies of grubs. Ecol. Lett. 4, 292–294.
- Veneklaas E J, Stevens J, Cawthray G R, Turner S, Grigg A M and Lambers H 2003 Chickpea and white lupin rhizosphere carboxylates vary with soil properties and enhance phosphorus uptake. Plant Soil 248, 187–197.
- Vessey J K, Pawlowski K and Bergman B 2005 N<sub>2</sub>-fixing symbiosis: legumes, actinorhizal plants, and cycads. Plant Soil 274, 51–78.
- Visser E J W and Voesenek L A C J 2005 Flooding of root systems – signals and signal-transduction pathways that lead to acclimation. Plant Soil 274, 197–214.
- Voesenek L A C J and Blom C W P M 1999 Stimulated shoot elongation: a mechanism of semi-aquatic plants to avoid submergence stress. *In* Plant Responses to Environmental Stresses: From Phytohormones to Genome Reorganization. Ed. H R Lerner. pp. 431–448. Marcel Dekker Inc, New York.
- Voesenek L A C J, Rijnders J H G M, Peeters A J M, van der Steeg H M and de Kroon H 2004 Plant hormones regulate fast shoot elongation under water; from genes to communities. Ecology 85, 16–27.
- Willey R W 1979 Intercropping its importance and research needs. Part 1. Competition and yield advantages. Field Crop Abstr. 32, 1–10.

- Wu H, Haig T, Pratley J, Lemerle D and An M 2000a Allelochemicals in wheat (*Triticum aestivum* L.): variation of phenolic acids in root tissues. J. Agric. Food Chem. 48, 5321–5325.
- Wu H, Pratley J, Lemerle D and Haig T 2000b Evaluation of seedling allelopathy in 453 wheat (*Triticuin aestivum*) accessions against annual ryegrass (*Lolium rigidum*) by the equalcompartment-agar method. Aust. J. Agric. Res. 51, 937–944.
- Yoder C K and Nowak R S 1999 Hydraulic lift among native plant species in the Mojave Desert. Plant Soil 215, 93–102.
- Zeuthen T and Klaerke D A 1999 Transport of water and glycerol in aquaporin 3 is gated by H<sup>+</sup>. J. Biol. Chem. 274, 21631–21636.
- Zhang F, Shen J, Li L and Lia X 2004 An overview of rhizosphere processes related with plant nutrition in major cropping systems in China. Plant Soil 260, 89–99.
- Zheng S J, Ma J F and Matsumoto H 1998 High aluminum resistance in buckwheat. I. Al-induced specific secretion of oxalic acid from toot tips. Plant Physiol. 117, 745–751.
- Zuo Y, Zhang F, Li X and Cao Y 2000 Studies on the improvement in iron nutrition of peanut by intercropping with maize on a calcareous soil. Plant Soil 220, 13–25.

Hans Lambers & Timothy D. Colmer School of Plant Biology Faculty of Natural and Agricultural Sciences The University of Western Australia 35 Stirling Highway, 6009, Crawley WA, Australia